

Comparing the effect of Seven isolated *Bacillus thuringiensis* against The Indian mealmoth (*Plodia interpunctella*), infesting during storage

Sabbour Magda¹ and Maysa E. Moharam²

1. Pests and Plant Prot., Dept . National Research Centre, Cairo Egypt
El-Tahrir St. Dokki, Cairo, Egypt

sabbourm@yahoo.com or magasabbour@gmail.com

2. Microbial Chemistry Department, Genetic Engineering & Biotechnology Div.

Abstract : Seven bacterial isolated strains, *Bacillus thuringiensis* B.T Dendrolimus , B.t thuringiensis , Bt Sotto 4A/4B , BT IP thurizide, Bt Toloworthi, Bt HD 210 and Bt HD 128 tested against the Indian meal moth (*Plodia interpunctella*, the LC₅₀ of the different bacterial strains , 103X10⁴, 98 X10⁴, 44 X10⁴, 40 X10⁴, 42 X10⁴, 110 X10⁴ and 177 X10⁴ after treated with different concentrations of bacterial strains, *Bacillus thuringiensis* B.T Dendrolimus , B.t thuringiensis , Bt Sotto 4A/4B , BT IP thurizide, Bt Toloworthi, Bt HD 210 and Bt HD 128,.

The effect of the number of eggs laid/ female of (*Plodia interpunctella*) were significantly decreased to 10.1±1.6 after treated with Bt Toloworthi as compared to 98.4±4.9 in the control after 120 days of the storage . The percentage of adult emergence significantly decreased to 10 after Bt Toloworthi treatments after 120 days as compared to 99% in the control. .

Keywords . Indian meal moth, (*Plodia interpunctella*), *Bacillus thuringiensis*

1. Introduction

The Indian meal moth, *Plodia interpunctella* (Hübner), is a very common household pest, feeding principally on stored food products. In fact, it has been called the most important pest of stored products that is commonly found in the home or in grocery stores in the Egypt. The larvae are general feeders, as they can be found in grain products, seeds, dried fruit, dog food, and spices Sabbour, 2003[1]. [2,3,4,5, 6] used the nanoparticles against the stored product insect pests, they found that the infections were significantly decreased when treated with the nanoparticles.

[7&8] found that, under laboratory conditions, the LC₅₀s, were significantly decreased when the adult female of grasshopper *Heteracris littoralis* treated with nano-destruxin and reached to 153X10⁴ spores/ml. Under semi field condition, the LC₅₀s of newly hatched nymphs, last nymphal stage and adult stages, 210 X 10⁴, 227 X 10⁴ and 224 X 10⁴ spores/ml [8].

[9] Lisansky suggested that the cutinophilic properties of the oil could allow a greater number of fungal conidia to penetrate the mouth parts of insects. Oil carriers can also distribute the inoculum over the thin intersegmental membranes, which are more rapidly penetrated by entomopathogenous fungi [9]. In addition, [10] found that the fungus *Beauveria bassiana* (Bals.-Criv.) Vuill. (Deuteromycotina: Hyphomycetes) killed the insect pests through the cuticle and it was not needed to be consumed by them. It is also mentioned

The present work aimed to explore the protective potency of

2. Material and methods

2.1. Rearing the Insect Pests

The target insect pests *Plodia interpunctella* was reared under laboratory conditions 28 ± 2°C and 60 ± 5% R.H on semi artificial diet (fine wheat with some endosperm), with 20% glycerin and 5% yeast powder. Groups of 100 one-day old eggs were placed each in 15 cm petridishes comprising a thin layer of diet. All cultures and experiments were held at 26 ± 2 °C and 70-80% R.H. with 16 hours light and 8 hours dark.

2.2. Microorganisms:

Bacillus thuringiensis B.T Dendrolimus B.t thuringiensis Bt Sotto 4A/4B BT IP thurizide Bt Toloworthi Bt HD 210 and Bt HD 128, were used in this study. The bacterial cultures were maintained on nutrient agar slants at 4°C.

2.3. Bacterial culture media:

The conventional laboratory culture broth, Nutrient broth, was used for culture preparation by mixing 5g peptone and 3g beef extract/ 1 L dist water. 50 ml of sterile medium was inoculated with one loopful of bacterial strain and incubated under shaking growth conditions on an orbital rotary shaker (125rpm) at 30°C for 72h.

2.4. Effect of the Microbial Control Agents: Isolated *Bacillus thuringiensis (Bt) B.T Dendrolimus, B.t thuringiensis, Bt Sotto, 4A/4B, BT IP thurizide, Bt Toloworthi, Bt HD 210 and Bt HD 128*; were used to test their activities on stored insect pests *B. incarnatus* adult beetles. The dead larvae of *B. incarnatus* were collected from the colony. The pathogen were isolated according to Salama et al [24]. The of Bt the tested concentrations were (500, 250, 125, 63, 32 and 16 ug/ml) (w/v). The rice pots were sprayed by tested concentrations of fungi or Bt and left to dry under laboratory conditions. Control treatment was made by feeding the larvae on untreated rice. The percentages of mortality were counted and calculated according to 50 [17], while LC50 were calculated through probit analysis according to [18]. The experiments were carried under laboratory conditions; 26 ± 2o C and 60- 70% R.H.

2.5. Effect of Storage Period on Weight Loss: To determine the impact of storage period on weight loss in the studied cultivars, samples of seeds were tested and as previously mentioned above during storage and weight loss was calculated according to Harris and Lindblad : Weight loss % = $\frac{(wu \times nd) - (wd \times nu)}{Wu (nd + nu)} \times 10$

Where:

Wd= weight of damaged seeds

nu= number of undamaged seeds

wu= weight of undamaged seeds

nd= number of damaged seeds

Data were subjected to analysis of variance (ANOVA) and means were compared by a least significant different test.

Results and Discussions

Table 1 show that the LC50 of the different bacterial strains, 103X10⁴, 98 X10⁴, 44 X10⁴, 40 X10⁴, 42 X10⁴, 110 X10⁴ and 177 X10⁴ after treated with different concentrations of bacterial strains, *Bacillus thuringiensis B.T Dendrolimus, B.t thuringiensis, Bt Sotto 4A/4B, BT IP thurizide, Bt Toloworthi, Bt HD 210 and Bt HD 128*,.

Table 2 show the effect of the number of eggs laid/ female of *P. incarnatus* were significantly decreased to 10.1±1.6 after treated with *Bt Toloworthi* as compared to 98.4±4.9 in the control after 120 days of the storage. The percentage of adult emergence significantly decreased to 10 after *Bt Toloworthi treatments after 120 days as compared to 99% in the control (Table2)*.

The effect of the different bacterial strains on seeds infections show that after different bacterial strains, the percentage of seeds infestation at the end of experiment were, 1, 4, 2, 1, 2, 15 and 20% after *Bacillus thuringiensis B.T Dendrolimus, B.t thuringiensis, Bt Sotto 4A/4B, BT IP thurizide, Bt Toloworthi, Bt HD 210 and Bt HD 128*, treatments as compared to 98 in the control (Table3).

Figure 1 show that beans seeds infestations were significantly decreased after all the bacterial strains treatments as compared to the control under store conditions.

The same results obtained by [15,16, 17,18,19,20,21,22,23, 24] applied different doses of the essential oils *Acorus calamus* to seeds of green gram *Vigna radiata* to protect them against *Callosobruchus chinensis* (L.) (Coleoptera: Bruchidae) and found that 1 ml/kg offered a high degree of protection up to a period of 135 days. Prolonged protection of the seeds was mainly due to a high adult mortality besides reduced oviposition and low hatching. [16] reported that foam sprayed with clove oil (5%) and placed between sacks caused the highest mortality. [25] reported that edible oils are potential control agents against *P. interpunctella* and play an important role in stored-grain protection. [15] mentioned that clove and eucalyptus oil vapours impaired the fecundity of bruchid beetles. Data proved promising oviposition deterrence toxicity and suppression of eggs and adult emergence. The effect of tested microbial control agents vapours on the reproduction of *P. interpunctella* was studied using the no choice test [5,6,7,8]. The reproduction of the weevils was reduced by the treatments with *B. bassiana*, followed by *M. anisopliae* and *B. thuringiensis*. Weevils laid eggs on treated seeds with *B. bassiana* but the number of eggs is always lower in treated seeds than in the control. [25] reported that edible oils are potential control agents against *P. interpunctella* and play an important role in stored-grain protection. [16] mentioned that clove and eucalyptus oil vapours impaired the fecundity of bruchid beetles. Data proved promising oviposition deterrence toxicity and suppression of eggs and adult emergence. [9], recorded that the LD50 for some formulations of *B. bassiana* was reduced It was suggested that the cutinophilic properties of the oil could allow a greater number of fungal conidia to penetrate the mouth parts of insects. Oil carriers can also distribute the inoculum over the thin intersegmental membranes, which are more readily penetrated by entomopathogenic fungi [9]. The increase in the pathogenicity of *B. bassiana* combined with mustard oil to *C. maculatus* beetles may be attributed to some degradation occurring at the structural level of the integument, which could have facilitated the penetration of the cuticle by the germ tube of the fungus. Similar results were obtained by [26] in *Manduca sexta* treated with *M. anisopliae* and the chitin-synthesis inhibitor dimilin. Synergistic effects of a combined application of *B. bassiana* and the chloronicotinyl insecticide imidacloprid on *Diaprepes abbreviatus* L. (Coleoptera: Curculionidae) were reported by [27]. Similar results obtained by [28,29]. In this respect, [23] applied different doses of essential oils of *Acorus calamus* seeds of green beans to protect them against pest infestation. Also, [21] reported that foam sprayed with clove oil (5%) and placed between sacks caused the highest mortality to *C. maculatus*. Similar results obtained by [30, 31, 32], and Similar results were found by [28, 29, 33, 34]. We choose gunny bags for further experiments due to their resistance compared to all other packing materials. The usage of the nano material were studied by [35] who used the nano chitosan and controlled the soya beans insects pests. Also, [36] who suggested that the application of the bioinsecticides which affected on decreasing the infestation, the number of infestations of *O. nubilalis*, *C. agamemnon* and *Sesamia cretica* significantly decreased; [37] Using of entomopathogenic fungi due to reduction the number of eggs laid / female after being treated with *B. brongniartii* and *N. rileyi* as compared the control. The emerged adults were decreased and the yield weight of potatoes increased in plots treated with *B. brongniartii* and *N. rileyi*. The yields weight of potatoes were significantly in plots treated with *B. brongniartii* and *N. rileyi* as compared in the control during seasons 2013 & 2014. [38] When *T. confusum* treated with the nano imidacloprid corresponding concentrations, the mortality percentage were significantly decreased to 70, 65 and 49 as compared to 2, 2 and 2 in the control. The mean number of the eggs laid /female of *T. castaneum* significantly decreased to when treated with imidacloprid and nano imidacloprid to 118.5 ± 2.1 and 18.6 ± 3.1 as compared to 289.9 ± 3.2 eggs/ female in the control. Larvae of *T. confusum* was more susceptible to the treatments than *T. castaneum* larvae, Nano-DE was more effective than natural-DE. The fecundity of tested insects was highly affected with both DE and nano-DE. The egg production was highly suppressed by nano-DE under stored conditions [39].

Acknowledgment. This project is supported by the National Research Centre / project no. (1020601) from 2013 – 2016.

4. Conclusion

Using of the bacteria *B. thuringiensis* against (*Plodia interpunctella*) showed significantly decrease of the insect pest infestations especially under store conditions.

5. References

- [1] Sabbour, M. M. 2003. The combined effects of some microbial control agents mixed with botanical extracts on some stored product insects. Pakistan. J. of Biol. Sci. 6 (1): 51-56.
- [2] Sabbour M.M. 2012. Entomotoxicity assay of two Nanoparticle Materials 1-(Al₂O₃ and TiO₂) Against *Sitophilus oryzae* Under Laboratory and Store Conditions in Egypt. Journal of Novel Applied Sciences. 1-4/103-108
- [3] Sabbour M.M. 2013. Entomotoxicity assay of Nanoparticle 4-(silica gel Cab-O-Sil-750, silica gel Cab-O-Sil-500) Against *Sitophilus oryzae* Under Laboratory and Store Conditions in Egypt. Sci. Re s. Rep. Vol., 1 (2), 67-74, 2013
- [4] Sabbour M.M. 2013. Entomotoxicity assay of Nano-particle 3-(Zinc oxide ZnO) Against *Sitophilus oryzae* Under Laboratory and Store Conditions in Egypt. Sci. Re s. Rep. Vol., 1 (2), 50-57, 2013

- [5] Sabbour M.M. 2013. Entomotoxicity assay of two Nanoparticle Materials 4a-(Al₂O₃ and TiO₂) Against *Sitophilus oryzae* Under Laboratory and Store Conditions in Egypt. Journal of Novel Applied Sciences. Sci. Res. Rep. Vol., 1 (2), 58-66, 2013.
- [6] Sabbour, M.M. 2014. Evaluating Toxicity of nano-Extracted Destruxin from *Metarhizium anisopliae* Against the grasshopper *Heteracris littoralis* in Egypt. J. Egypt. Acad. Environ. Develop. 15(2): 1-7.
- [7] Sabbour, M.M. 2014. Evaluating toxicity of extracted nano -Destruxin against the desert locust *Schistocerca gregaria* in Egypt. J. Egypt. Acad. Environ. Develop. 15(2): 9-17.
- [8] Lisansky, S. 1989: Biopesticides fall short of market projections. Performance Chem. 16: 387-396.
- [9] Abd El-Gawad, H. & Abd El-Aziz, A. 2004: Evaluation of different integrated pest management concept for controlling the legumes beetles *Callosobruchus maculatus* (F.) and *Callosobruchus chinensis* (L.) on faba bean and cowpea seeds. Bull. Entomol. Egypt. Entomol. Econ. 30: 105-122.
- [10] Leiderer, P. and Dekorsy, T. (2008). Interactions of nanoparticles and surfaces Tag der m Ä undlichen Pr Äufung: 25. April. <http://www.ub.unikonstanz.de/kops/volltexte/2008/5387/>;
- [11] Abbott, W. S. 1925: A method of computing the effectiveness of an insecticide. J. Econ. Entomol. 18: 265-267.
- [12] Lwande W., Hassanalli A., Njoroge P.W., Bentley M.D., DelleMonache F., Jondiks J.I. 1985. A new 6a-hydroxy petrocarpan with insect antifeedant and antifungal properties from the root of *Tephrosia hildebrandtu*. Vatl. Ins. Sci. Appl. 6 (4): 537-541.
- [13] Sokal R.R. and Rohlf F.J. [1981]. The Principles and Practice of Statistics in Biological Research. Freeman. San Francisco, p. 859.
- [14] Batta, Y. A. 2004: Control of the rice weevil (*Sitophilus oryzae* L., Coleoptera: Curculionidae) with various formulations of *Metarhizium anisopliae*. Crop Prot. 23: 103-108.
- [15] Deshpande, R. S., Adhikary, P. S. & Tipris, N. P. 1974: Stored grain pest control agents from *Nigella sativa* and *Pogostemon heyneanus*. Bull. Grain Technol. 12: 232-234.
- [16] Jacobson, M. 1975: Insecticides from plants. A review of literature. USDA Agriculture Handbook 461: 1954-1971.
- [17] Baby, J. K. 1994: Repellent and phago deterrent activity of *Sphaeranthus indicus* extract against *Callosobruchus chinensis*. In: Highley, E., Wright, E. J., Banks, H. J., Champ, B. R. (eds.): Proceeding of the 6th International Working Conference on Stored-Product Protection, Canberra, 17-23 April 1994, CAB International, Wallingford, UK: 746-748.
- [18] Rodriguez, E. & Levin, D. H. 1975: Biochemical Parallelism of Repellents and Attractants in Higher Plants and Arthropods. In: Wallace, J. M., Mansell, R. L. (eds.): Recent advances in phytochemistry biochemical interaction between plants and insects. Plenum Press, New York: 215-270.
- [19] Abd El-Aziz, S. E. & Ismail, I. A. 2000: The effectiveness of certain plant oils as protections of broad bean against the infestation by *Bruchus incaratus*. Schm. (Coleoptera: Bruchidae) during storage. Ann. Agric. Sci. 45: 717-725.
- [20] Abd El-Aziz, S. E. 2001: Persistence of some plant oils against the bruchid beetle, *Callosobruchus maculatus* (F.) (Coleoptera: Bruchidae) during storage. Arab. Univ. J. Agric. Sci. 9: 423-432.
- [21] Sabbour, M. M. 2002: Evaluation studies of some bio-control agents against corn borer in Egypt. Ann. Agr. Sci. 47: 1033-1043.
- [22] Chander, H. & Ahmed, S. M. 1986: Efficacy of oils from medicinal plants as protectants of green gram against the plus beetle. *Callosobruchus chinensis*. Entomon 11: 21-28.
- [23] Ketoh, G. K., Koumaglo, H. K., Glitho, I. A. and Huignard, J. 2006: Comparative effects of *Cymbopogon schoenanthus* essential oil & piperitone on *Callosobruchus maculatus* development. Fitoterapia 77: 506-510.
- [24] Shaaya, E., Kostjukovski, M., Eilberg, J. & Sukprakarn, C. 1997: Plant oils as fumigants and contact insecticides for the control of stored-product insects. J. Stored Prod Res. 33: 7-15.
- [25] Hassan, A. E. M. & Charnley, K. A. 1989: Ultrastructural study of penetration by *Manduca sexta*. J. Invertebr. Pathol. 54: 117-124.
- [26] Quintela, E. D. & McCoy, C. W. 1998: Synergistic effect of imidacloprid and two entomopathogenic fungi on the behaviour and survival of larvae of *Diaprepes abbreviatus* (Coleoptera: Curculionidae) in soil. J. Econ. Entomol. 91: 110-122.
- [27] Sabbour, M. M. & Abd El Aziz, S. E. 2007: Evaluation of some bioinsecticides and packing materials for protecting broad bean against *Callosobruchus maculatus* (Coleoptera: Bruchidae) infestation during storage. In: Proceedings of the 2nd International Conference of Economic entomology, Cairo, Egypt, 8-11 December, 2007, Entomological Society of Egypt: 255-267.
- [28] Sabbour, M. M. & Abd El-Aziz, S. E. 2010: Efficacy of some bioinsecticides against *Bruchidius incarnatus* (Boh.) (Coleoptera: Bruchidae). Infestation during storage. J. Plant Prot. Res. 50: 28-34.
- [29] Saxena, B. P., Koul, O & Tikku, K. 1976: Non-toxic protectant against the stored grain insect pests. Bull. Grain Technol. 14: 190-193.
- [30] Surabaya, S., Babu, C. K., Krishnappa, C. & Murty, K. C. K. 1994: Use of locally available plant products against *Callosobruchus chinensis* in red gram. Mysore J. Agric. Sci. 28: 325-345.

- [31] Maheshwari, H. K, Sharma, M. K. & Dwivedi, S. C. 1998: Effectiveness of repelin as surface protectant against plus beetle, *Callosobruchus chinensis* infesting cowpea. Int. J. Trop. Agric. 16: 229-232.
- [32] Leelavathi, K., Rao, P. H, Indrani, D. & Shurpalekar, S. R. 1984: Physico-chemical changes in whole wheat flour (Atta) and resultant atta during storage. J. Food Sci. Technol. 21: 23-27.
- [33] Upadhyay, R. K., Thangaraj, M and Jaiswal, P. K. 1994: Storage studies of suji in different packages. J. Food Sci. Technol. 31: 494-496. Vassilakos, T. N., Athanassiou, C. G., Kavallieratos, N. G. and Vayias, B. J. 2006: Influence of temperature on the insecticidal effect of *Beauveria bassiana* in combination with diatomaceous earth against *Rhyzopertha dominica* and *Sitophilus oryzae* on stored wheat. Biol. Contr. 38: 270-281.
- [34] Sahab, A. F.; Waly, A.I., Sabbour, M. M. and Lubna S. Nawar. 2015. Synthesis, antifungal and insecticidal potential of Chitosan (CS)-g-poly (acrylic acid) (PAA) nanoparticles against some seed borne fungi and insects of soybean. Vol.8, No.2, pp 589-598.
- [35] Sabbour M.M and S.M. Singer. 2015. Efficacy of Nano *Isaria fumosorosea* and *Metarhizium flavoviride* against Corn Pests under Laboratory and Field Conditions in Egypt. International Journal of Science and Research (IJSR). ISSN (Online): 2319-7064.
- [36] Sabbour, Magdaand MA Abdel-Raheem. 2015. Determination the efficacy of *Beauveria brongniartii* and *Nomuraea rileyi* against the potato tuber moth *Phthorimaea operculella* (Zeller). American J. of innovative research and applied sci. 197-202.
- [37] Sabbour, M.M. 2015. Efficacy of some nano-Imidacloprid against red flour beetle *Tribolium castaneum* and confused flour beetle, *Tribolium confusum* (Coleoptera: Tenebrionidae) under laboratory and store conditions. Advances in Biochemistry & Biotechnology. 1-13.
- [38] Sabbour, M.M. and Shadia El-Sayed Abd-El-Aziz. 2015. Efficacy of some nano-diatomaceous earths against red flour beetle *Tribolium castaneum* and confused flour beetle, *Tribolium confusum* (Coleoptera: Tenebrionidae) under laboratory and store conditions. Bull. Env. Pharmacol. Life Sci., Vol 4 [7] June 2015: 54-59.

Table 1. Effect of the tested *B. thuringiensis* on *P. interpunctella*.

| Target pathogen | LC ₅₀ | S | V | 95% Confidence limits |
|--------------------------|-----------------------|-----|-----|-----------------------|
| <i>B.T Dendrolimus</i> | 103 X 10 ⁴ | 0.1 | 1.4 | 89-149 |
| <i>B.t thuringiensis</i> | 98 X 10 ⁴ | 2.0 | 1.3 | 77-133 |
| <i>Bt Sotto 4A/4B</i> | 44 X 10 ⁴ | 0.1 | 1.2 | 66-121 |
| <i>BT IP thurizide</i> | 40 X 10 ⁴ | 0.1 | 1.2 | 33-122 |
| <i>Bt Toloworthi</i> | 42 X 10 ⁴ | 0.1 | 1.1 | 35-124 |
| <i>Bt HD 210</i> | 110 X 10 ⁴ | 0.2 | 1.2 | 99-145 |
| <i>Bt HD 128</i> | 177 X 10 ⁴ | 1.1 | 1.3 | 57-189 |

Table 2. Effect of different bacterial strains treatments *Plodia interpunctella* under laboratory conditions.

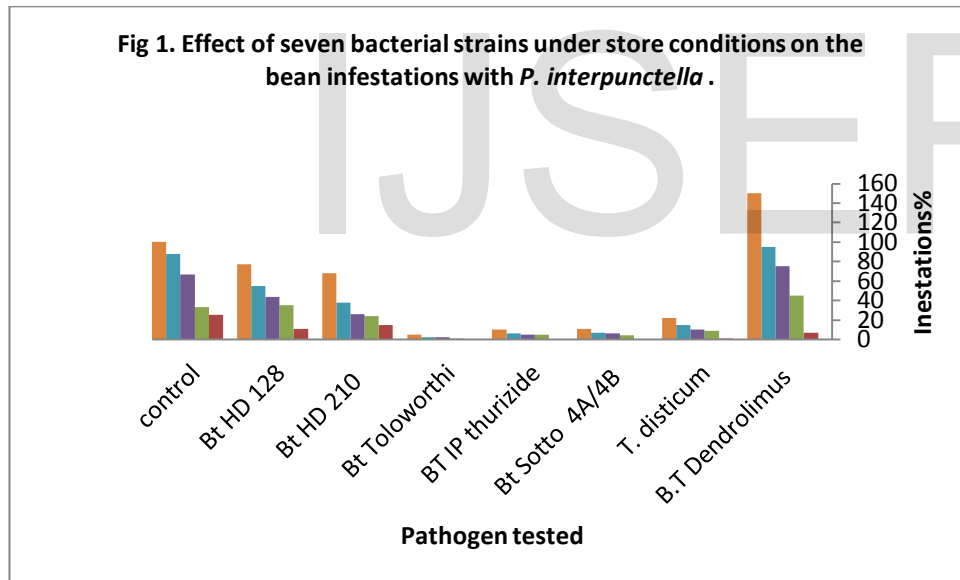
| Insects | Days Storage interval | | | | | | | |
|--------------------------|------------------------|------------------------------|------------------------|------------------------------|------------------------|------------------------------|------------------------|------------------------------|
| | 25 | | 45 | | 90 | | 120 | |
| | no. of eggs /♀±S.E. | % adult emergence (F1) | no. of eggs /♀±S.E. | % adult emergence (F1) | no. of eggs /♀±S.E. | % adult emergence (F1) | no. of eggs /♀±S.E. | % adult emergence (F1) |
| <i>B.T Dendrolimus</i> | 19.8±1.5 | 7 | 20.8±1.9 | 10 | 23.8±1.2 | 10 | 27.1±1.4 | 12 |
| <i>B.t thuringiensis</i> | 18.8±1.6 | 4 | 18.8±3.7 | 7 | 11.8±1.3 | 10 | 19.8±4.9 | 11 |
| <i>Bt Sotto 4A/4B</i> | 7.8±1.1 | 4 | 7.8±3.1 | 9 | 11.3±1.2 | 10 | 15.1±3.9 | 11 |
| <i>BT IP thurizide</i> | 9.7±5.9 | 6 | 7.8±2.8 | 8 | 11.3±5.1 | 8 | 12.1±1.4 | 10 |
| <i>Bt Toloworthi</i> | 6.8±1.5 | 5 | 6.4±6.9 | 7 | 8.1±1.7 | 8 | 10.1±1.6 | 10 |
| <i>Bt HD 210</i> | 22.6±1.4 | 55 | 21.8±1.4 | 26 | 28.8±1.9 | 58 | 28.9±1.7 | 60 |

| | | | | | | | | |
|------------------|----------|-----|----------|-----|----------|----|----------|----|
| <i>Bt</i> HD 128 | 38.4±5.1 | 60 | 25.8±1.8 | 29 | 29.8±6.9 | 59 | 29.1±8.9 | 69 |
| <i>Control</i> | 99.9±5.9 | 100 | 9.8±1.8 | 100 | 98.8±3.9 | 99 | 98.4±4.9 | 99 |

Table 3. The effect of the storage period on seed infestations after bacterial treatments during storage.

| Treatments | % of seeds infestations | | % of seeds wt loss | |
|--------------------------|-------------------------|--------------------|---------------------|--------------------|
| | Start of Experiment | End of experiments | Start of Experiment | End of experiments |
| <i>B.T Dendrolimus</i> | 0 | 1 | 0 | 6 |
| <i>B.t thuringiensis</i> | 0 | 4 | 0 | 2 |
| <i>Bt Sotto 4A/4B</i> | 0 | 2 | 0 | 0 |
| <i>BT IP thurizide</i> | 0 | 1 | 0 | 0 |
| <i>Bt Toloworthi</i> | 0 | 2 | 0 | 0 |
| <i>Bt</i> HD 210 | 0 | 15 | 0 | 20 |
| <i>Bt</i> HD 128 | 0 | 20 | 0 | 37 |
| <i>Control</i> | 5 | 98 | 0 | 87 |

Fig 1. Effect of seven bacterial strains under store conditions on the bean infestations with *P. interpunctella*.



IJSER